



SVENSKA GRUPPEN AML

Familial Leukaemia — what genetics is teaching us Jude Fitzgibbon

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What's in Store

- Run through the current landscape of familial leukaemia.
- Genetic testing
- The transcription factor CEBPA
- Identify new loci
- New ideas emerging from the study of inherited leukaemia.

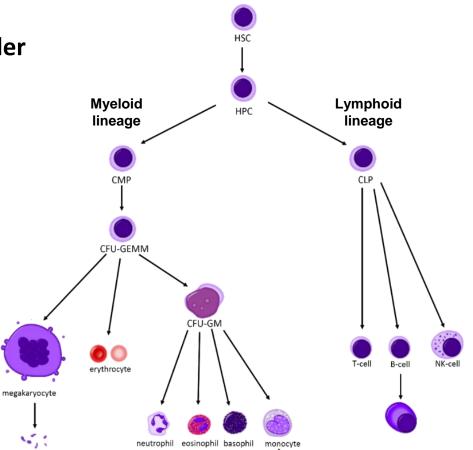
Acute Myeloid Leukaemia (AML)

- Haematopoeitic Stem Cell Disorder

- **Median age:** 68 years

- **Incidence:** 3/100,000 in Europe

- 5-year overall survival: 42%



Familial AML/MDS

- > 95 % of all AML cases are sporadic.
- << 5% of the cases where two or more affected individuals are found within the same family.
- New WHO Entity 2016 provisional diagnostic category for heritable myeloid malignancies



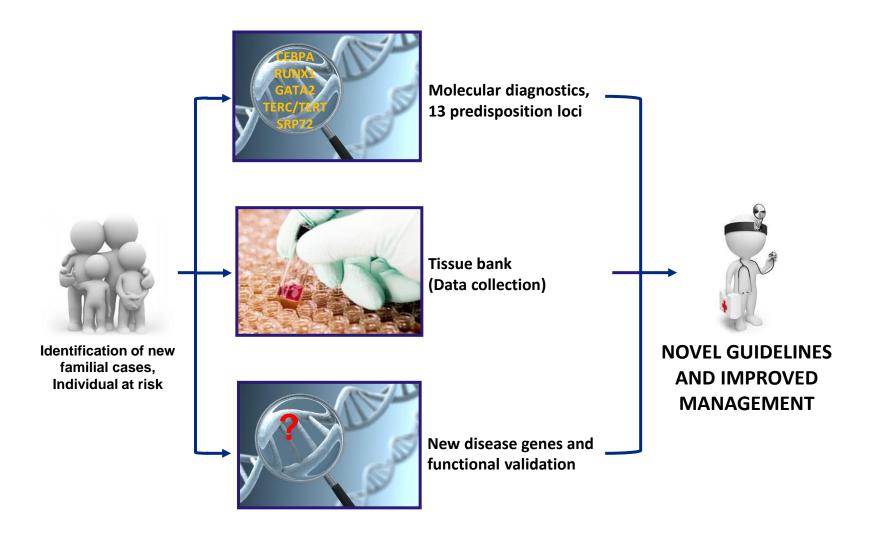
Clinical challenges in Familial AML

- 1. Recognition of inherited forms of these diseases is difficult;
 - Patients may be unaware of their predisposition.
 - Wide variation in the age of onset and disease phenotype.
 - Absence of customized diagnostics.
- 2. Variable penetrance of the disease mutations: symptomatic and asymptomatic carriers.

3. How to manage patients with GL mutation? - Paucity of clinical guidelines.



Roadmap to improve diagnosis, treatment and management



Familial MDS/AML Genetic Discovery

Familial MDS/AML can be divided into three groups:

- Examples where approved testing exists.
- Emerging from basic research and requiring validation.
- Without an identified genetic basis.

(Little is known regarding secondary genetic mutations)

Current Mutational Landscape

GENE	AUTHORS	YEAR
RUNX1	Song et al. Boston, Massachusetts, USA	1999
TERC	Vulliamy et al. London, UK	2001
CEBPA	Smith <i>et al.</i> London, UK	2004
TERT	Hiroki Yamaguchi et al. Atlanta, USA	2005
GATA2	Hahn <i>et al</i> . Adelaide, Australia	2011
ANKRD26	Noris <i>et al.</i> Pavia, Italy	2011
SRP72	Kirwan <i>et al.</i> London, UK	2012
ACD	Guo <i>et al.</i> Philadelphia, USA	2014
ETV6	Zhang <i>et al.</i> Seattle, Washington, USA and Leila Noetzli <i>et al.</i> Colorado USA	2014/2015
ATG2B/GSKIP	Saliba <i>et al.</i> Villejuif, France	2015
DDX41	Polprasert <i>et al</i> . Cleveland, USA	2015
SAMD9	Narumi <i>et al</i> . Tokyo, Japan	2016
SAMD9L	Tesi <i>et al.</i> Stockholm, Sweden	2017

Current Mutational Landscape – 13 genes and counting

Myeloid malignancies only

AML: CEBPA

MDS/AML: DDX41

MPNs/AML: ATG2B/GSKIP

Cytopenias and/or platelet dysfunction

FPD/AML: *RUNX1 GATA2* deficiency

Thrombocytopenia 2: ANKRD26

Thrombocytopenia 5: ETV6

Bone marrow failure syndromes

Telomere syndromes: TERT, TERC, ACD

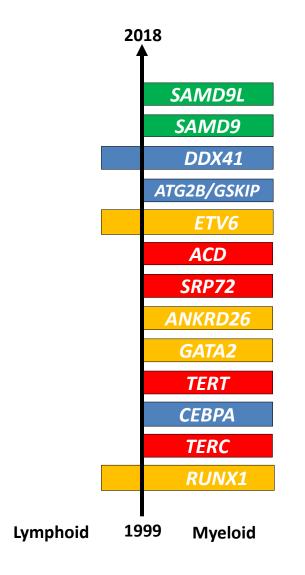
Aplastic anemia/MDS: SRP72

Other syndromes

Cytopenia, immunodeficiency, MDS, and neurological

symptoms: SAMD9L

MIRAGE syndrome: SAMD9



Targeted sequencing panel for familial MDS/AML

Index cases were tested for mutation in 11 known disease genes

West Midlands Regional Genetics Laboratories

Birmingham Women's NHS Foundation Trust

ACD, ANKRD26, ATG2B, CEBPA, DDX41, ETV6, GATA2, RUNX1, SRP72, TERC and TERT



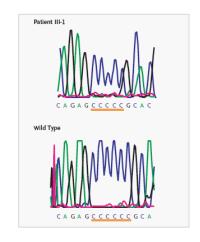
Our Interest in Familial CEBPA

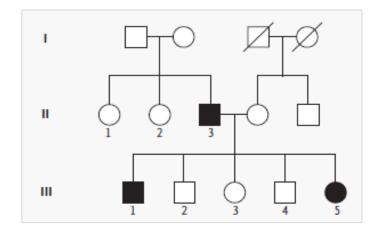


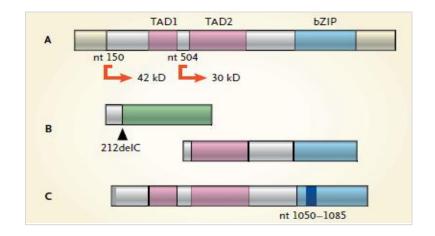
Mutation of CEBPA in Familial Acute Myeloid Leukemia

BRIEF REPORT

Matthew L. Smith, M.B., B.S., Jamie D. Cavenagh, M.D., T. Andrew Lister, M.D., and Jude Fitzgibbon, Ph.D.







International collaboration on Familial *CEBPA*-AML: 25 patients – 11 pedigrees

Smith *NEJM*, 2004 (UK)

Sellick Leukemia, 2005 (UK)

Pabst et al, JCO, 2008 (Switzerland)

Renneville Leukemia, 2009 (France)

Nanri GCC, 2010 (Japan)

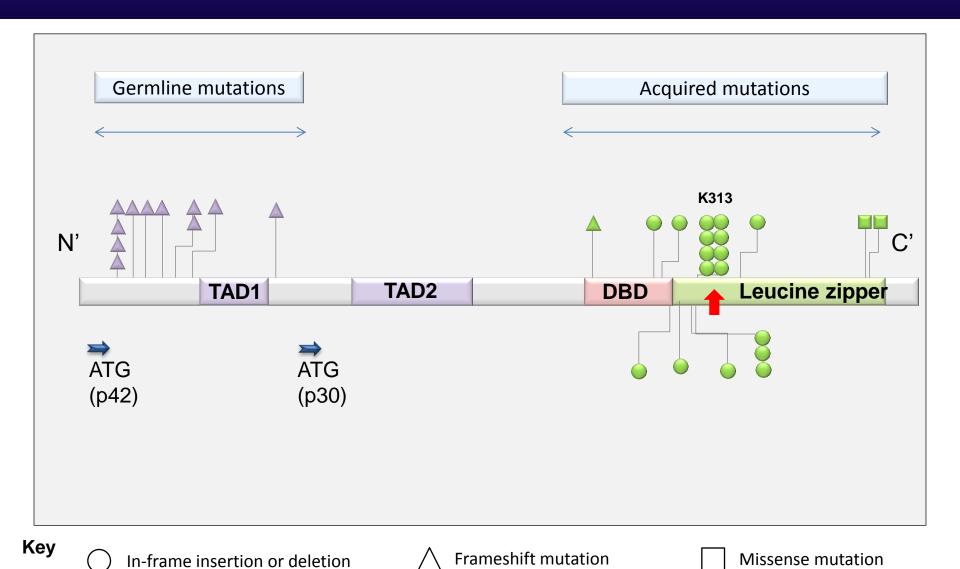
Stelljes *Leukemia*, 2011 (Germany)

Taskesen Leukemia, 2011 (Germany)

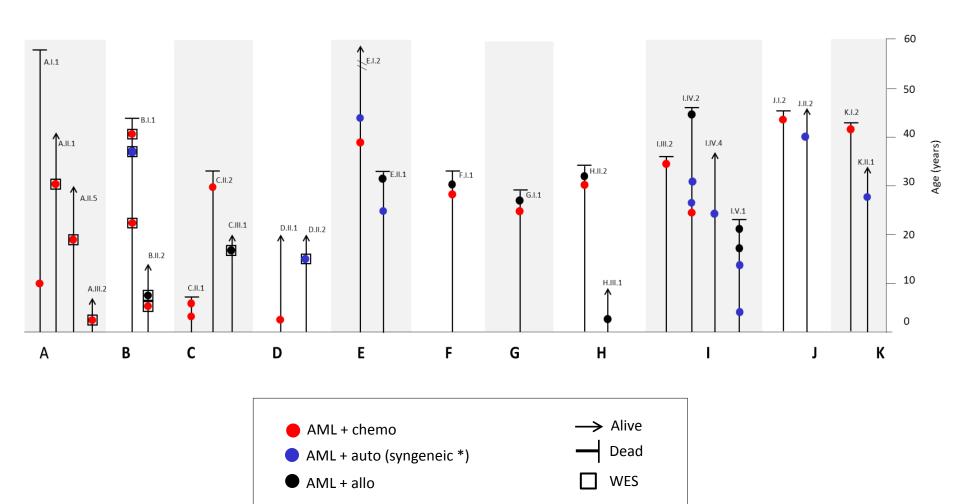
Savic A, unpublished (Serbia)

Debeljak *Haematologica* 2013 (Slovenia)

Distribution of CEBPA mutations

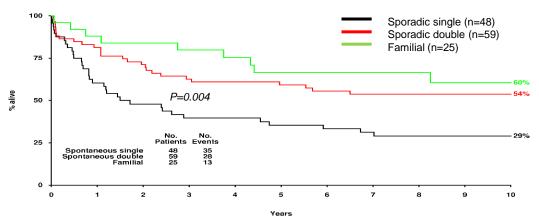


Timeline of clinical events in all 25 Familial *CEBPA* AMLs

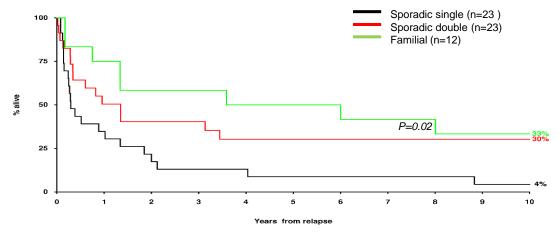


Comparison Familial versus MRC CEBPA series

A) Overall survival

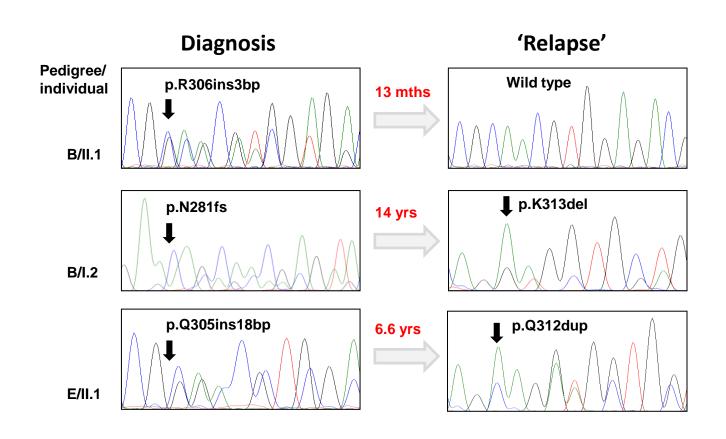


B) Post-relapse survival

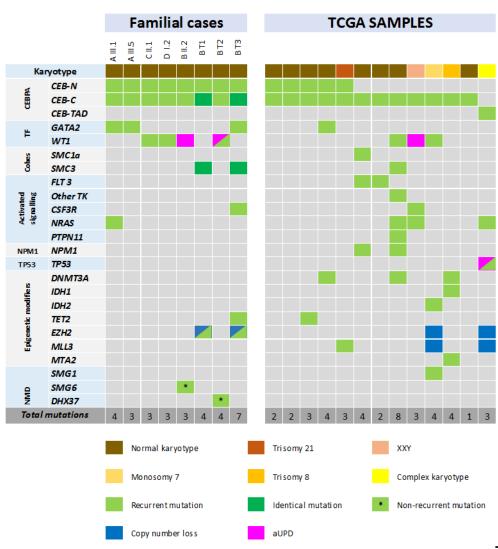


Patients are cured of initial disease but are predisposed to new leukemic episodes

New CEBPA C-terminal mutations at relapse



Familial CEBPA reminiscent of dmCEBPA

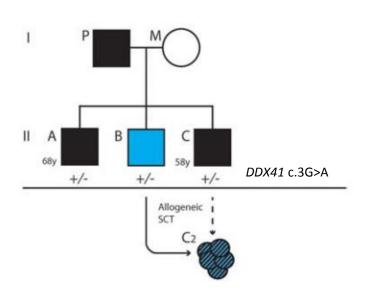


Collective experience

- Chemotherapy alone can be used to treat patients without adverse disease features.
- High frequency of late 'chemosensitive' relapses, where intensive consolidation, preferably with allogeneic transplantation, may well prove beneficial in preventing further disease events.
- Perform germ-line testing in sporadic CEBPA AML of <40 yrs at diagnosis.
- Counselling of affected individuals and asymptomatic carriers requires full knowledge and understanding of the implications of inherited *CEBPA* mutations and, above all, recognition and consideration for an individual's choice.
- Where possible, we advocate screening of all potential sibling donors, preparing for both treatment escalation in affected cases and identifying asymptomatic mutation carriers for counselling and surveillance.

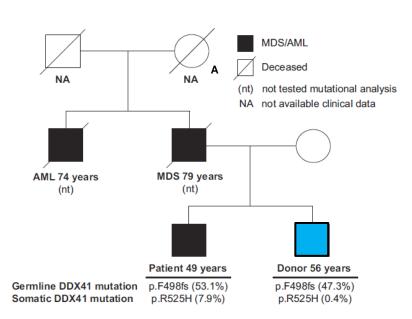
Donor cell leukaemia in *DDX41* families

FAMILY 1



Berger et al, 2017

FAMILY 2

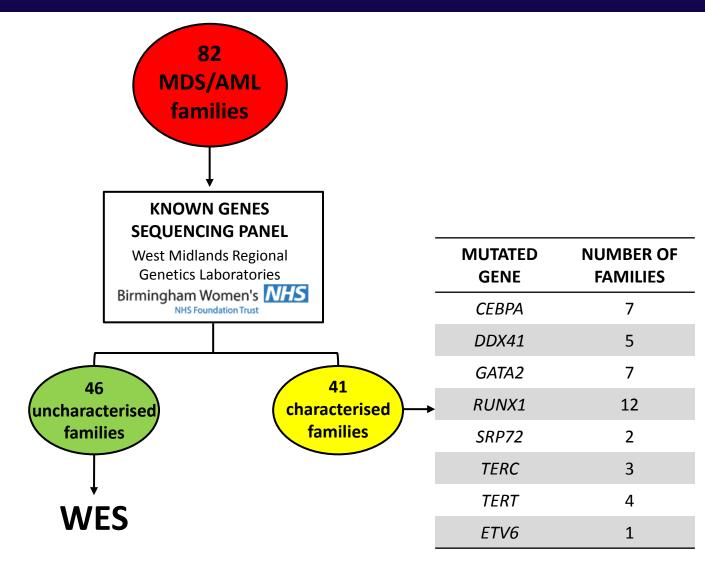


Kobayashi et al, 2017

Lesson from familial leukaemia

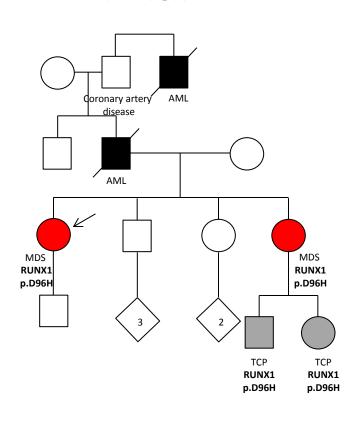
- Genes not mutated in sporadic AML Novel mechanism
- Under appreciated mechanisms of mutation
- Clustering of mutations within pedigrees Host genetics
- A clever way of making a monosomy
- Disease Latency and penetrance Protection factor

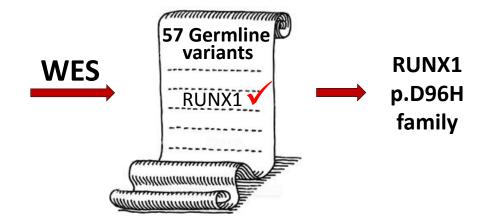
Our cohort of MDS/AML families – new Loci (Vulliamy, Dokal & Fitzgibbon)



Challenges in identifying a familial MDS/AML loci

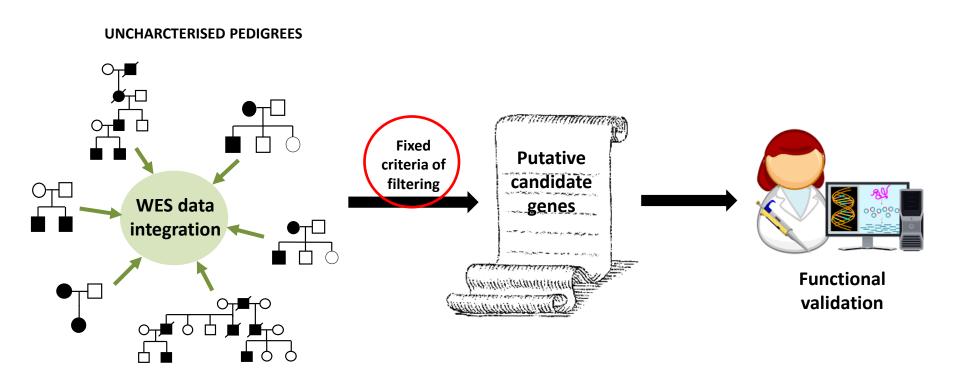
PEDIGREE 1





Challenges in identifying a familial MDS/AML loci

FAMILIES WITHOUT A CLEAR CANDIDATE



Challenges in identifying a familial MDS/AML loci

HOW TO FIX CRITERIA OF FILTERING?

- 1. Variant frequency (e.g. ExAC < 0.0001)
- 2. Predicted to be damaging (*Polyphen, MutationTaster, SIFT, Provean* scores)
- 3. In at least 2 families (?)
- 4. Others: family phenotype, protein function, type of mutation, mutated in sporadic AML, other scores (PhyloP, GERP, and CADD)...

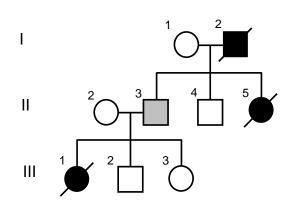
Some putative candidate genes

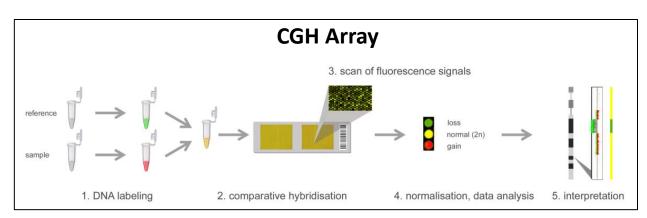
GENE	VARIANT FREQUENCY (ExAC)	PREDICTED TO BE DAMAGING	IN AT LEAST 2 FAMILIES	PROTEIN FUNCTION
ADA	0	YES	YES (2 families)	Adenosine deaminase (SCID related)
DHX34	< 0.0001	YES	YES (4 families)	DExH-box helicase (involved in NMD)
PDCD4	0	YES	YES (2 families)	Potential TSG (downregulated in AML)
PRR23B	0	Not all variants	YES (3 families)	Proline Rich 23B (regulated by <i>GATA2</i>)
VPS13C	< 0.0001	Not all variants	YES (5 families)	Vacuolar Protein (mitochondrial function)

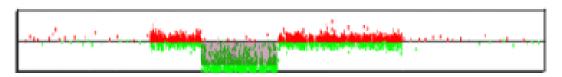
Lessons from familial leukaemia

- Genes not mutated in sporadic AML Novel mechanism
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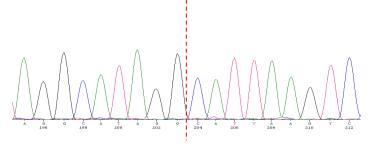
Atypical mutations: intragenic deletion in *RUNX1* – 20% of pedigrees







RUNX1 - Chr 21- 1.21Mb



Deletion breakpoints chr21:36389492-37056053 - 66,6561 bp deletion

Germ-line GATA2 – synonymous mutations

Major classes of germline mutations in cases *GATA2* deficiency:

- 1. Truncating (stop-gain, frameshift, splice-site);
- 2. Missense within zinc fingers;
- 3. Noncoding substitutions in the conserved EBOX-GATA-ETS regulatory region (intron 4);
- 4. In frame indels and whole gene deletions.
- 5. Silent (synonymous) exonic substitutions Splice defects

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2449 Systematic Assessment of GATA2 Genetic Variation Reveals the Presence of Novel Disease–Causing Synonymous Exonic Mutations

Bone Marrow Failure

Program: Oral and Poster Abstracts

Session: 508. Bone Marrow Failure: Poster II

Sunday, December 10, 2017, 6:00 PM-8:00 PM

Bldg A, LvI 1, Hall A2 (Georgia World Congress Center)

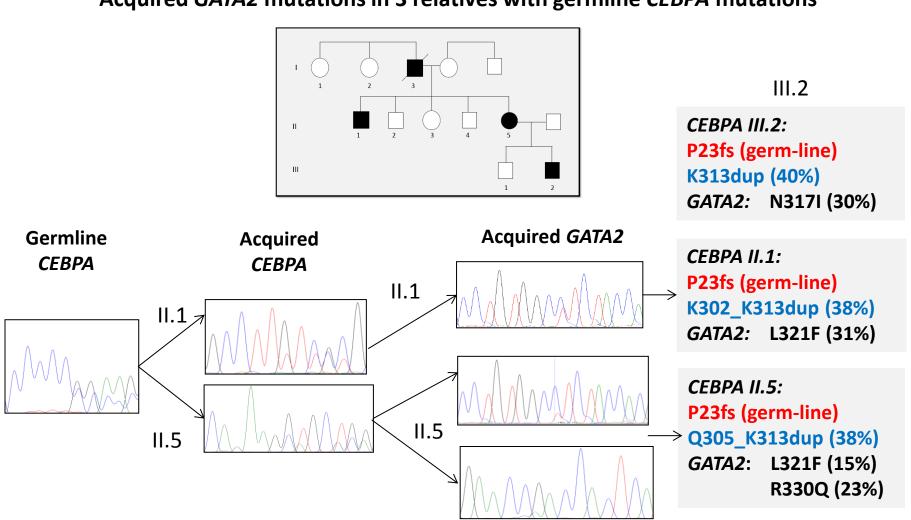
Emilia J. Kozyra, MSc<sup>1,2*</sup>, Victor Pastor Loyola, MSc<sup>3,4*</sup>, Claudia Wehr<sup>5,6*</sup>, Sushree Sangita Sahoo, M.Sc.<sup>3,7,8*</sup>, Rebecca Voss<sup>9*</sup>, Enikoe Amina Szvetnik<sup>10*</sup>, Shinsuke Hirabayashi, MD<sup>11*</sup>, Albert Catala<sup>12*</sup>, Henrik Hasle, MD, PhD<sup>13,14</sup>, Marry M. Van den Heuvel–Eibrink, MD<sup>15,16</sup>, Krisztián Kallay, MD<sup>17*</sup>, Riccardo Masetti, MD, PhD<sup>18*</sup>, Barbara De Moerloose, MD, PhD<sup>19*</sup>, Markus Schmugge, MD<sup>20</sup>, Owen Smith, MD<sup>21*</sup>, Marek Ussowicz<sup>22*</sup>, Jan Stary<sup>23*</sup>, Ester Mejstrikova<sup>24</sup>, Ramunė Pasaulienė<sup>25*</sup>, Irith Baumann, Dr. med.<sup>26*</sup>, Gudrun Göhring, MD<sup>27*</sup>, Brigitte Schlegelberger, Prof. Dr.<sup>27</sup>, Ulrich Salzer<sup>5,28*</sup>, Michael Lübbert<sup>29</sup>, Eirini Trompouki, PhD<sup>30*</sup>, Charlotte M. Niemeyer, MD<sup>31</sup> and Marcin W. Wlodarski. MD. PhD<sup>32</sup>
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Lessons from familial leukaemia

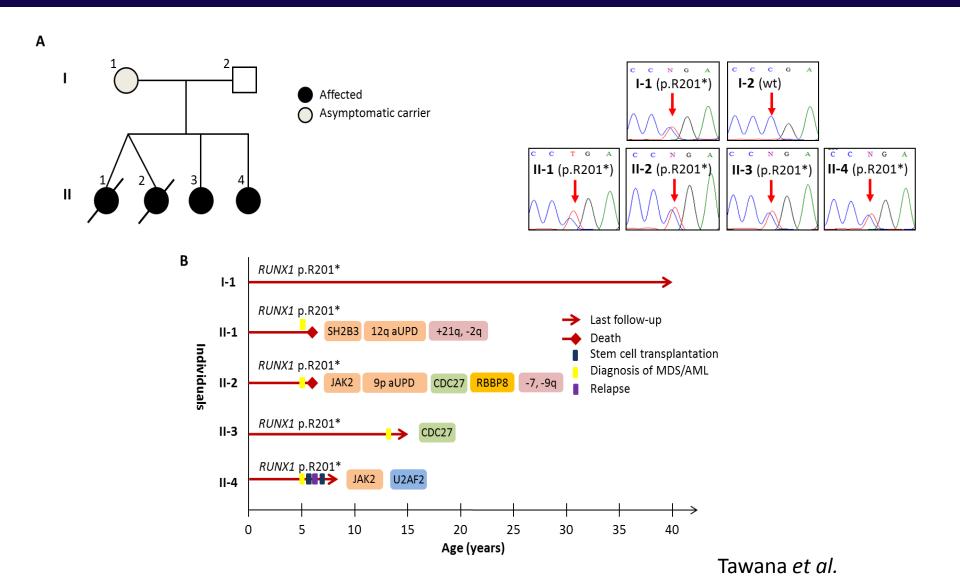
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Clustering of same secondary mutations in pedigrees - *GATA2*

Acquired GATA2 mutations in 3 relatives with germline CEBPA mutations



Disease Latency - RUNX1

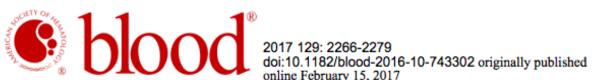


Lesson from familial leukaemia

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SAMD9L Mutations and Loss of Chromosome 7





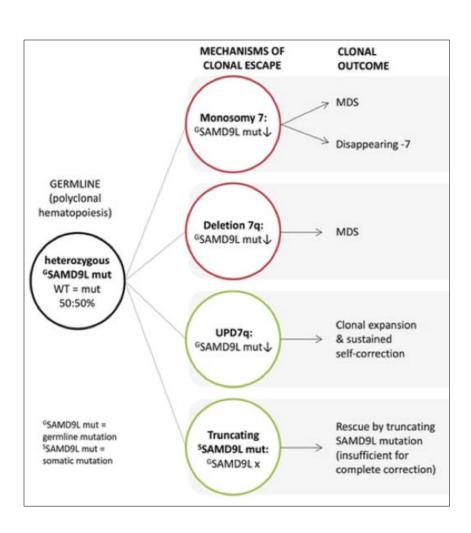
Gain-of-function SAMD9L mutations cause a syndrome of cytopenia, immunodeficiency, MDS, and neurological symptoms

Bianca Tesi, Josef Davidsson, Matthias Voss, Elisa Rahikkala, Tim D. Holmes, Samuel C. C. Chiang, Jonna Komulainen-Ebrahim, Sorina Gorcenco, Alexandra Rundberg Nilsson, Tim Ripperger, Hannaleena Kokkonen, David Bryder, Thoas Fioretos, Jan-Inge Henter, Merja Möttönen, Riitta Niinimäki, Lars Nilsson, Cornelis Jan Pronk, Andreas Puschmann, Hong Qian, Johanna Uusimaa, Jukka Moilanen, Ulf Tedgård, Jörg Cammenga and Yenan T. Bryceson

- 1. Heterozygous germline gain-of-function (GoF) mutations in SAMD9L are associated with cytopenia, immunodeficiency, MDS and neurological disease (e.g. ataxiapancytopenia).
- 2. These **GoF** mutations cause decreased cell proliferation compared to WT protein and in turn, lead to loss of SAMD9L-mutant allele and monosomy 7.

So there is a section to lose the mutation (loss of Chromosome 7) to increase proliferation.

Familial MDS and Transient Monosomy 7 as a Sole Clinical Manifestation of *SAMD9L*-related Disease



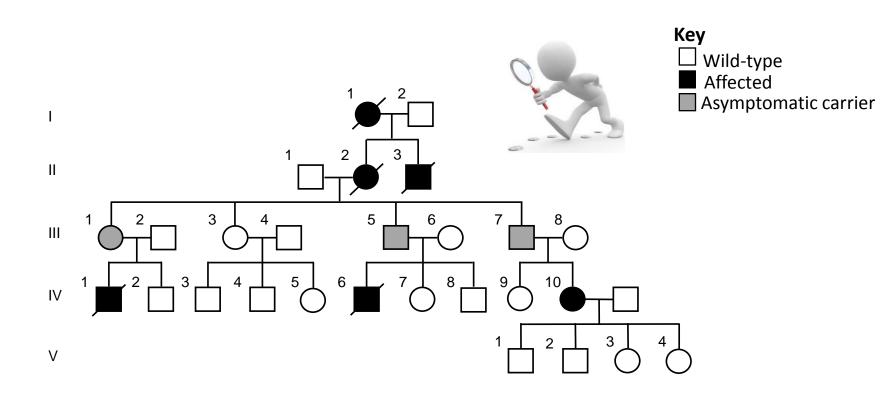


Constitutional SAMD9L mutations cause familial myelodysplastic syndrome and transient monosomy 7

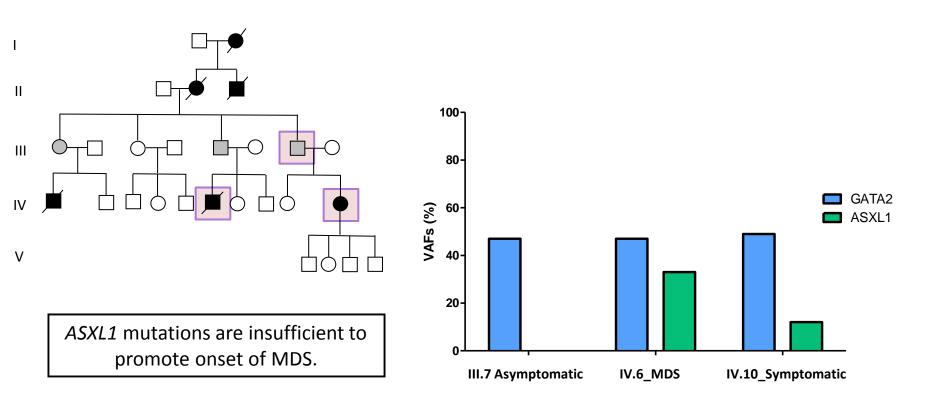
by Victor B. Pastor, Sushree Sahoo, Jessica Boklan, Georg C. Schwabe, Ebru Saribeyoglu, Brigitte Strahm, Dirk Lebrecht, Matthias Voss, Yenan T. Bryceson, Miriam Erlacher, Gerhard Ehninger, Marena Niewisch, Brigitte Schlegelberger, Irith Baumann, John C. Achermann, Akiko Shimamura, Jochen Hochrein, Ulf Tedgård, Lars Nilsson, Henrik Hasle, Melanie Boerries, Hauke Busch, Charlotte M. Niemeyer, and Marcin W. Wlodarski

- Non-random loss of mutant SAMD9L allele can be achieved by:
 - 1. Complete (Monosomy 7)
 - 2. Partial (deletion 7q)
 - 3. UPD 7q
 - 4. Somatic truncating *SAMD9L* mutations

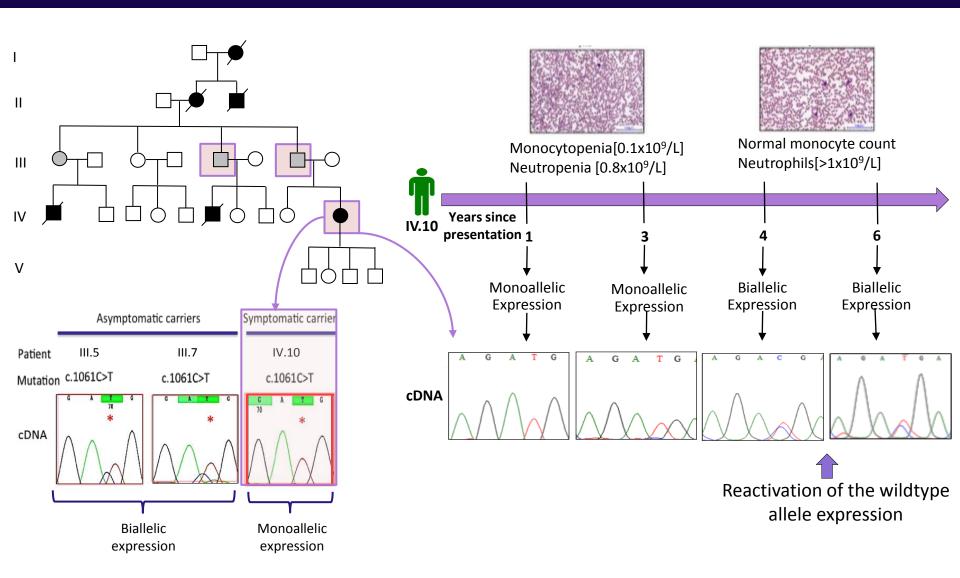
Reduced penetrance in *GATA2*-mutated MDS/AML - p.T354M mutations



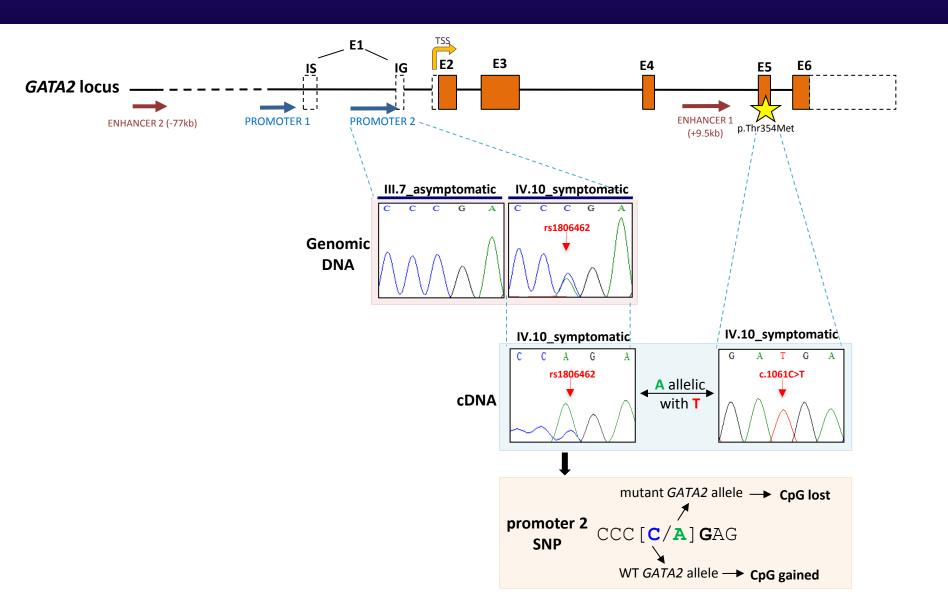
1. ASXL1 mutation (p.Gly646TrpfsTer12) as a secondary genetic event in GATA2 symptomatic carriers



2. GATA2 monoallelic expression discriminates between symptomatic and asymptomatic carriers and correlates with clinical parameters

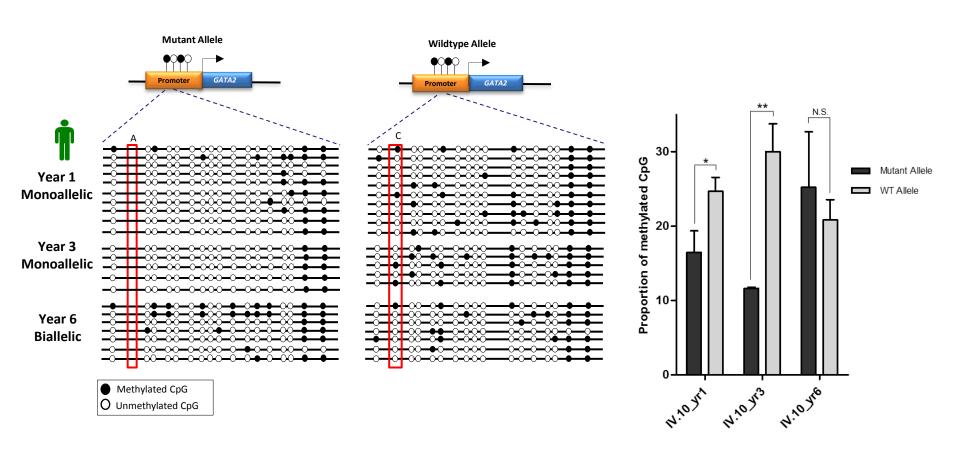


3. A CpG-SNP provided a means of distinguishing between alleles



4. Epigenetic reprogramming regulates monoallelic GATA2 expression

1. DNA methylation as a mechanism of silencing the WT allele.

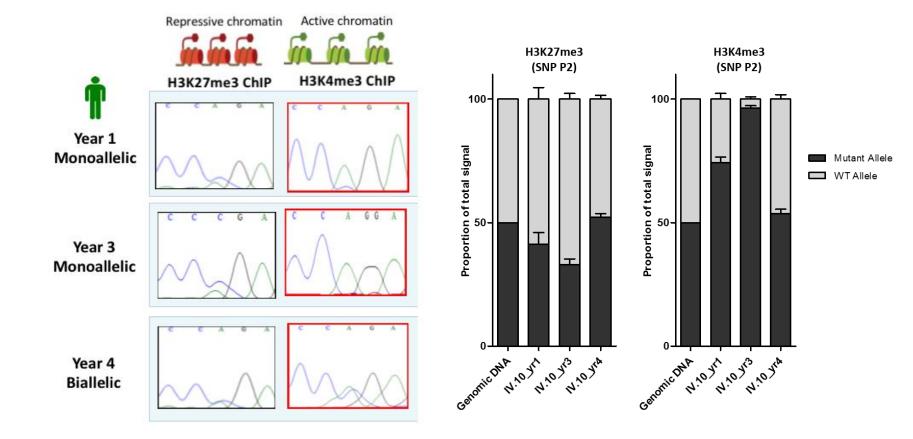


4. Epigenetic reprogramming regulates monoallelic GATA2 expression

1. DNA methylation as a mechanism of silencing the WT allele.

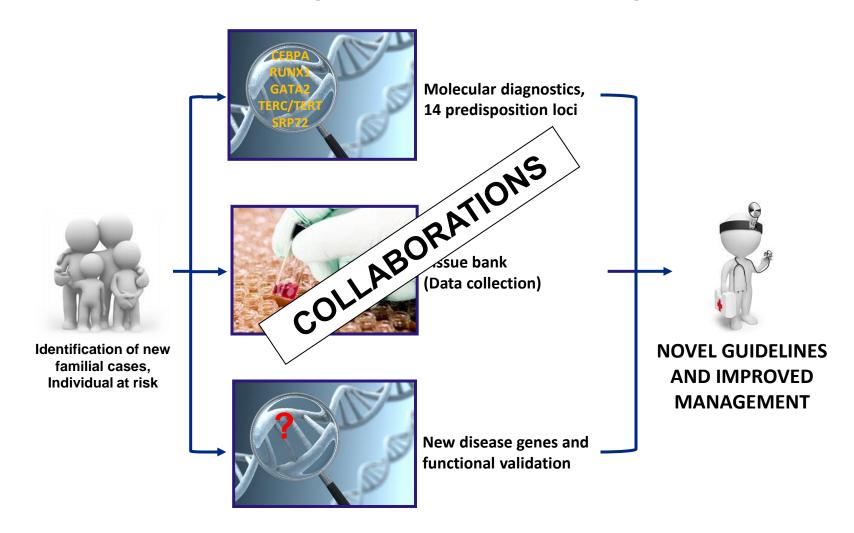
Mutually exclusive

2. Enhanced H3K4me3 promoter deposition on the mutant allele.

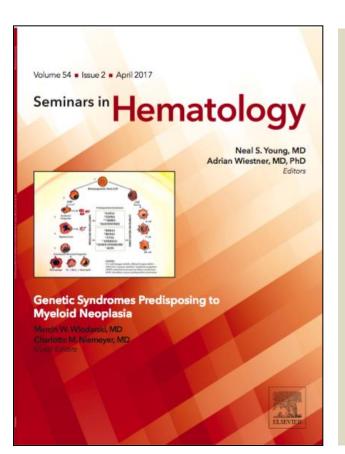


Collaboration is Key

How to improve diagnosis, treatment and management?



Seminars in Hematology – Special Issue 2017 Edited Marcin W. Wlodarski and Charlotte M. Niemeyer



- Recognition of familial myeloid neoplasia in adults
- Practical considerations for diagnosis and management of patients and carriers
- RUNX1 deficiency (familial platelet disorder with predisposition to myeloid leukemia, FPDMM)
- GATA2 deficiency and related myeloid neoplasms
- Familial CEBPA-mutated acute myeloid leukemia
- DDX41-related myeloid neoplasia
- ETV6 in hematopoiesis and leukemia predisposition
- Classical inherited bone marrow failure syndromes with high risk for myelodysplastic syndrome and acute myelogenous leukemia
- Cancer predisposition syndromes associated with myeloid malignancy

Acknowledgements

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Gavin Ryan Yvonne Wallis Susanna Akiki Joanne Mason

Csaba Bodor

Our Global Collaborators

Some Key Groups Worldwide

Hamish Scott (Australia)
Charlotte Niemeyer (Germany)
Inderjeet Dokal/Tom Vulliamy (UK)
Lucy Godley (USA)

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